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Diabetes mellitus worsens outcomes in tPA thrombolytic stroke therapy

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The demonstrated benefit of tissue type plasminogen activator (tPA) thrombolytic therapy has been a landmark achievement in stroke therapy. However, not all patients respond equally, and some patients have worse outcomes. In particular, diabetes mellitus is recognized as a clinically important vascular co-morbidity that leads to lower recanalization rates and increased risk of hemorrhagic transformation. In this mini-review, we summarize recent advances in the understanding of the underlying mechanisms involved in worse stroke outcome in diabetic patients. Potential pathological factors that relate to suboptimal recanalization include higher plasma plasminogen activator inhibitor-1 levels, diabetic atherogenic vascular damage, glycation of the tPA receptor annexin A2, alterations in fibrin clot density, and impaired collaterals. Factors that may contribute to hemorrhagic transformation include hyperglycemia, vascular oxidative stress, inflammation, tPA neurovascular toxicity, and dysfunction in extracellular proteolysis balance. A better understanding of these complex pathways may eventually lead to novel ways of counteracting the negative effects of diabetes in ischemic stroke.

Keywords: ischemic stroke, tissue-type plasminogen activator, diabetes mellitus, hyperglycemia, recanalization rate, intracerebral hemorrhage

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Tissue plasminogen activator (tPA): thrombolytic stroke therapy and its limitations

Ischemic stroke is a cerebrovascular event. Despite enormous research efforts that include many clinical trials, intravenous administration of recombinant tPA remains the only FDA-approved treatment for ischemic stroke, and the most beneficial proven intervention for emergency treatment of ischemic stroke (Chapman et al., 2014; Whiteley et al., 2014). tPA thrombolytic stroke therapy is based on the "recanalization hypothesis," which posits that reopening of occluded vessels by lysing the clot will improve clinical outcome in acute ischemic stroke (AIS) through regional reperfusion and salvaging threatened tissues (Whiteley et al., 2014). Recanalization is an important predictor of stroke outcome after thrombolysis. The demonstrated clinical benefit of thrombolytic therapy has been a landmark discovery in the treatment of AIS. However, there are disadvantages to the use of thrombolytic therapy including lower thrombolytic perfusion rate, short therapeutic treatment time window, and risk of hemorrhagic transformation (Alexandrov and Grotta, 2002; Rubiera et al., 2005; Bambauer et al., 2006; Weintraub, 2006). Overcoming these disadvantages and making tPA work more effectively is a priority that will aid in advancing AIS treatment (Davalos, 2005; Thomassen and Bakke, 2007). Although other thrombolytic agents are being tested, none have been shown to be an effective replacement for tPA (Adams et al., 2007). Importantly, exogenous tPA may worsen ischemia-induced blood brain-barrier (BBB) disruption, elevate the risk of symptomatic intracranial hemorrhage, and in part, reduce the therapeutic time window (Chapman et al., 2014). Recent clinical investigations have indicated potential opportunities to improve tPA therapy. For instance, European ECASS III trial showed that intravenous tPA given up to 4.5 hr after symptom onset improved clinical outcomes (Cronin, 2010). In the Echoplanar Imaging Thrombolytic Evaluation trial, thrombolysis 4.5-6 hr after stroke onset reduced infarct growth and increased the rate of reperfusion, which was associated with good neurological and functional outcome (Picanco et al., 2014). Moreover, recent trials point to potential avenues to improve patient access by imaging-based patient selection, and to facilitate rapid and complete reperfusion of the penumbra (Manning et al., 2014). One such avenue includes endovascular thrombectomy performed from 6 to 24 hrs after stroke onset (Nogueira et al., 2018). This review will only focus on intravenous tPA thrombolytic reperfusion therapy (IV tPA).

Although imaging-based patient selection may help to identify more ischemic stroke patients who are candidates for IV tPA, IV tPA is still associated with increased intracranial hemorrhagic transformation, which remains the most threatened complication for thrombolytic stroke therapy (Harsany et al., 2014). There are a number of risk factors associated with tPA stroke therapy-mediated intracerebral hemorrhagic transformation, including diabetes mellitus (DM), post-stroke hyperglycemia, older age, larger infarct, and high blood pressure (Miller et al., 2011; Faigle et al., 2014; Shrestha et al., 2014). In this mini review, we focus on the role and mechanism(s) of DM as a risk factor for the lower recanalization rate and intracerebral hemorrhagic transformation after tPA thrombolytic stroke therapy. Although the molecular mechanisms underlying DM-related complications after stroke remain to be further elucidated, a better understanding of these complex pathways may eventually lead to novel ways of counteracting the negative effects of diabetes in stroke (Fan et al., 2014; Shrestha et al., 2014; Li et al., 2017).

DM in stroke clinical epidemiology

DM is a major risk factor for cardiovascular diseases, including stroke. Globally, stroke mortality rates have fallen, but stroke incidence and its sequelae have significantly increased over

the last three decades (Hill, 2014). Diabetes is a recognized independent risk factor for stroke and is associated with higher morbidity and mortality rates (Chen et al., 2016). Clinical epidemiological investigations have documented that diabetic patients are 2 to 6 times more susceptible to ischemic stroke; about 30% of stroke patients are diabetic and more than 90% of them comprise type 2 diabetes mellitus (T2DM) (Beckman et al., 2013). Additionally, other cardiovascular metabolic risk factors including obesity, dyslipidaemia, and hypertension are often comorbid with T2DM, and may in concert contribute to higher stroke risks when compared to patients with similar risk profiles without diabetes (Chen et al., 2016). Clinically T2DM stroke patients have nearly double the mortality rate and worse neurological outcomes compared to non-diabetic stroke patients (Air and Kissela, 2007; Tureyen et al., 2011). Importantly T2DM stroke patients respond less favorably to tPA therapy due to lower recanalization rates (Molina et al., 2001; Molina et al., 2004; Tang et al., 2016), but have higher risks of hemorrhagic transformation (Linfante et al., 2002; Kwon et al., 2004).

DM: an unfavorable comorbidity for tPA thrombolytic stroke therapy

1. DM is a risk factor for lower recanalization rates after tPA thrombolysis

Intravenous thrombolysis (IVT) with recombinant tPA is a proven beneficial treatment for AIS when given within 4.5 hr of symptom onset (Jauch et al., 2013). Although patients with AIS and diabetes can achieve substantial benefit from IVT (Reiter et al., 2014), several large clinical trials examining ischemic stroke patients treated with IVT reported associations between DM and/or post-stroke hyperglycemia and unfavorable neurological outcome, hemorrhagic transformation, and death (Linfante et al., 2002; Kwon et al., 2004). Other clinical studies also showed that DM or post-stroke hyperglycemia was associated with lower recanalization rates in IVT treated stroke patients (Molina et al., 2001; Molina et al., 2004; Tang et al., 2016). These clinical studies suggested that there is an impaired fibrinolytic response in the setting of DM and/or hyperglycemia. Although the detailed mechanisms underlying DM-related complications after stroke remain to be defined, we speculate that the following pathological factors might contribute to the reperfusion resistance or lower recanalization rate after IVT in DM stroke patients.

The first pathological factor that we hypothesize may underlie lower recanalization rates in stroke patients with DM is higher plasma plasminogen activator inhibitor 1 (PAI-1) levels. Hypercoagulative status in DM is mainly attributed to elevated platelet activation and higher circulating PAI-1 levels, which might be partially responsible for the lower recanalization rate of IVT (Pandolfi et al., 2001; Vaidyula et al., 2006; Lemkes et al., 2010; Tjarnlund-Wolf et al., 2012). It has been shown that diabetes and metabolic syndrome are associated with increased plasma PAI-1 levels in patients at risk of atherothrombosis (Alessi et al., 2007; Aso, 2007). Biologically, PAI-1, the main and potent endogenous t-PA inhibitor, could potentially affect the therapeutic efficacy of exogenously administered t-PA by inhibiting its actions. Although clinical investigation is largely lacking, a few experimental and clinical reports suggest that a fibrinolytic profile upon admission is associated with symptomatic hemorrhagic transformation and recanalization resistance in stroke patients treated with t-PA reperfusion therapy (Ribo et al., 2004a; Ribo et al., 2004b; Montaner, 2009; Walter et al., 2010). It has been shown that lower baseline plasma PAI-1 and thrombin-activable fibrinolysis inhibitor (TAFI) levels predict symptomatic hemorrhagic transformation (Ribo et al., 2004b), whereas higher plasma PAI-1 levels upon admission predict t-PA thrombolytic resistance (Ribo et al., 2004a). However, we should be cautious when using

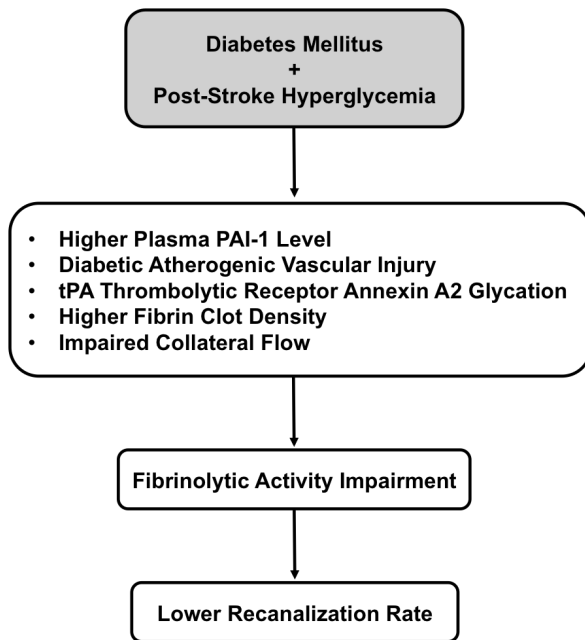


Figure 1. A schematic outline to link pathological association between diabetes mellitus plus post stroke hyperglycemia and lower recanalization rate after tPA thrombolytic stroke therapy. The potential pathological factors that underlying this therapeutic shortcoming include higher plasma PAI-1 level, diabetic atherogenic vascular injury, glycation of tPA thrombolytic receptor annexin A2, higher fibrin clot density, and impaired collateral flow. These factors all contribute to fibrinolytic activity impairment in exogenous tPA that clinically presents as a lower recanalization rate.

PAI-1 levels as a predictive indicator. In addition, other risk factors, such as atherothrombosis and pre-existing vascular comorbidities in most patients with stroke also need to be considered (Whiteley, 2011). Thus the role of circulating PAI-1 levels in the variable reperfusion rate and hemorrhagic transformation risk following t-PA thrombolytic stroke therapy needs to be further elucidated (Tjarnlund-Wolf et al., 2012).

Another possible pathological factor that may be involved in the lower recanalization rate in DM stroke patients treated with IVT is diabetic atherogenic vascular damage. Systemic inflammation and atherogenic pathological processes in the cerebral vascular walls are present in DM, resulting in elevation of vascular inflammation, BBB integrity disruption, impaired vascular fibrinolysis, and eNOS activity (Lenart et al., 2016). Once ischemic stroke occurs under diabetic state, there may be more robust platelet activation, fibrin deposition, circulating inflammatory cell accumulation, and new thrombosis formation at the occluded vascular injury site, which might lead to thrombolytic resistance (Ly et al., 2017; Venkat et al., 2017).

One more potential pathological factor underlying the low recanalization rate with DM is glycation of tPA thrombolytic receptor annexin A2. Annexin A2 is a fibrinolytic receptor of tPA that acts to accelerate tPA-converted plasmin generation (Ling et al., 2004). Our previous studies showed recombinant annexin A2 (rA2) in combination with low-dose tPA improved thrombolytic efficacy and long-term neurological outcomes after embolic focal ischemia in rats (Wang et al., 2014; Jiang et al., 2015; Fan et al., 2017). Impaired fibrinolysis on the surface of endothelial cells has been identified as a key pathological factor in thrombotic vascular complications in patients with diabetes (Alzahrani and Ajjan, 2010). An immunochemical and biochemical study examining endothelial plasma membrane proteins that are glycosylated in diabetes, has identified annexin

A2 as one of the three major glycosylated proteins labeled by the anti-glycitolysine (Ghitescu et al., 2001). In cultured human brain microvascular endothelial cells, we found hyperglycemia for 7 days significantly reduced cell surface fibrinolytic activity, and also decreased tPA, plasminogen, and annexin A2 mRNA and protein expressions, while increasing PAI-1 levels. Hyperglycemia significantly increased advanced glycation end products (AGE)-modified forms of total cellular and membrane annexin A2. The hyperglycemia-associated reduction in fibrinolytic activity was fully restored upon incubation with recombinant annexin A2 (rA2). However, neither the hyperglycemia induced increases in AGE-modified annexin A2 nor exogenous tPA were reversed, supporting the hypothesis that hyperglycemia causes dysfunction in the endothelial membrane protein annexin A2, thereby leading to an overall reduction in fibrinolytic activity (Dai et al., 2013). Our experimental findings also support the previous speculation that impaired fibrinolysis due to glycation of endothelial annexin A2 may be partially responsible for worse neurological outcome and resistance to tPA reperfusion in diabetic stroke complications, as an acquired annexinopathy (Gugliucci and Ghitescu, 2002).

Additionally, a higher fibrin clot density may also be considered a pathological factor for the low recanalization rate in stroke patients with DM. It has been reported that DM patients exhibit increased maximal fibrin clot strength (Maatman et al., 2018) and increased fiber density (Dunn et al., 2005). Prolonged duration of T2DM is associated with a pro-thrombotic fibrin clot phenotype (Konieczynska et al., 2014). These observations suggest a denser fibrin clot in DM patients might also contribute to the increase in tPA thrombolytic resistance (Dunn et al., 2006; Tang et al., 2016). Lastly, impaired collateral flow compensation may also underlie the low recanalization rate in stroke patients with DM (Kimura et al., 2009; Liebeskind et al., 2014). Cerebral vasoreactivity and collateral circulation are important protective mechanisms against cerebral ischemia. Clinical investigations have suggested that more robust collateral grade was associated with better recanalization, reperfusion, and subsequently better clinical outcomes (Liebeskind et al., 2014). Experimental studies have documented impaired collateral flow compensation during chronic cerebral hypoperfusion and after focal ischemic stroke in mice with T2DM (Akamatsu et al., 2015; Nishijima et al., 2016), suggesting that the impaired collateral flow might not directly affect thrombolysis, but may contribute to the lower reperfusion efficacy of tPA thrombolytic therapy and worse neurological outcomes in DM stroke patients (Kimura et al., 2009; Tang et al., 2016).

2. DM is a risk factor for tPA-induced intracerebral hemorrhagic transformation

Symptomatic intracranial hemorrhage (SICH) is a devastating complication of intravenous thrombolysis treatment that is associated with high mortality (Seet and Rabinstein, 2012). SICH rates, reported to be from 3-7%, vary considerably between studies and these differences may relate to the differences in the criteria used to define SICH (Seet and Rabinstein, 2012; Reiter et al., 2014). There are a number of risk factors associated with tPA stroke therapy-mediated hemorrhagic transformation, including post-stroke hyperglycemia, older age, larger infarct, and high blood pressure (Miller et al., 2011; Faigle et al., 2014). Post-stroke hyperglycemia is present in all preexisting diabetes patients (about 37% of stroke patients), and 50% of non-diabetic stroke patients (Allport et al., 2006; Kruyt et al., 2010). History of DM, combined with the severity of post-stroke hyperglycemia, is associated with poor clinical outcome after stroke and thrombolysis (Ribo et al., 2005; Bruno et al., 2008; Poppe et al., 2009; Ahmed et al., 2010; Desilles et al., 2013). For example, in the NINDS r-tPA Stroke Trial,

in patients treated with tPA within 3 hours of onset, serum glucose level was an independent predictor that suggested direct correlation with symptomatic hemorrhagic transformation (Bruno et al., 2002). This was replicated in the PROACT II trial, where symptomatic hemorrhagic transformation occurred in 35% of patients with serum glucose values greater than 200 mg/dL (Kase et al., 2001). In another study using data from the prospective, multicenter Canadian Alteplase for Stroke Effectiveness Study (CASES), in the cohort of IV-tPA-treated stroke patients, admission hyperglycemia was independently associated with increased risk of death, hemorrhagic transformation, and poor functional status at 90 days (Poppe et al., 2009). Although evidence supports an increased risk of hemorrhage due to tPA in DM patients with hyperglycemia, the mechanisms underlying this effect remain to be fully elucidated (Ishrat et al., 2012; Fan et al., 2014; Whiteley et al., 2014; Kanazawa et al., 2017).

Interactions among multiple factors, including hyperglycemia-mediated vascular oxidative stress, neuroinflammation-mediated injury, ischemic insult, and tPA neurovascular toxicity contribute to the extracellular proteolysis dysfunction- BBB damage-intracerebral hemorrhagic transformation process (Lo et al., 2002; Wang and Lo, 2003; Wang et al., 2004; Won et al., 2011; Hafez et al., 2014). Hyperglycemia may cause vascular oxidative stress, which occurs very early after the onset of ischemia/reperfusion injury via overproduction of reactive oxygen species (ROS). Oxidative stress generated during stroke is a critical event leading to BBB disruption with secondary vasogenic edema and hemorrhagic transformation of infarcted brain tissue, restricting the benefit of thrombolytic reperfusion (Kaur et al., 2004; Wang et al., 2004). ROS can also directly oxidize and damage BBB structures. Furthermore, ROS is an upstream intermediate of pathophysiological mechanisms during reperfusion injury that links protease activation to vascular leakage (Gasche et al., 2001; Jian Liu and Rosenberg, 2005). The importance of oxidative stress in stroke and tPA thrombolytic-related vasculature disruption has been well-documented in several animal studies employing antioxidant plus tPA combination treatments in embolic stroke models (Asahi et al., 2000; Lapchak et al., 2001; Lapchak et al., 2002). In human stroke, increased oxidative stress and its direct relationship to matrix metalloproteinase (MMP)-9 expression was observed (Kelly et al., 2008), in accordance with preclinical animal studies.

Post-ischemic neuroinflammation-mediated BBB leakage after stroke is a progressive and interactive process, and largely depends on the activation, expression, and secretion of proinflammatory mediators (e.g. cytokines) from both cerebral and peripheral cells (Rosenberg, 2002; Gidday et al., 2005; Amantea et al., 2013). Oxidative stress is a major stimulator of inflammatory cytokine production and protease secretion by microglia, leukocytes, and brain resident cells of the neurovascular unit (Wang and Lo, 2003; Lee et al., 2004; Pun et al., 2009). Experimental investigations have suggested that neuroinflammation-mediated extracellular matrix proteolysis dysfunction is the key pathological mechanism contributing to BBB disruption after stroke, mainly by means of early elevated cytokines, release of leukocytes into circulation, adhesion of leukocytes to the injured cerebrovasculature, leukocyte brain infiltration, and release and activation of proteases (Simi et al., 2007; Najjar et al., 2013). Leukocyte-microvessel interactions, and subsequent infiltration of leukocytes into the ischemic brain play prominent roles in the development of secondary damage (Amantea et al., 2013), resulting in edema, microvascular permeabilization, and hemorrhage via secreted free radicals, cytokines/chemokines, lipid-derived mediators, and proteases (Wang and Lo, 2003; Fagan et al., 2004; Borlongan et al., 2012). It has been clearly recognized that proteases secreted

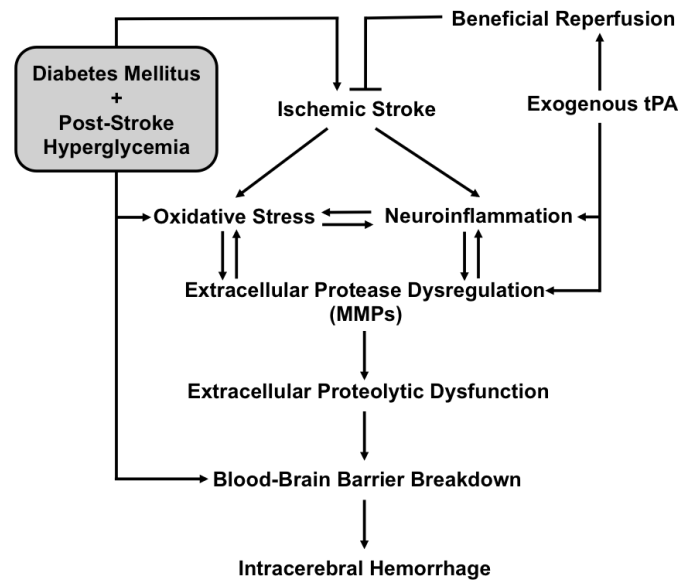


Figure 2. A schematic outline to link multiple interactions among pathological factor, which consequently mediates BBB breakdown and hemorrhagic transformation after tPA thrombolytic therapy in diabetes mellitus stroke patients. Properly administered exogenous tPA facilitates reperfusion of the ischemic brain and the rescue of compromised tissue. Exogenous tPA may also activate extracellular proteases and neuroinflammation to potentiate extracellular proteolysis dysfunction. Diabetes and post-stroke hyperglycemia directly prime cerebrovascular oxidative stress and inflammation. Ischemic stroke fuels early elevation in oxidative stress and neuroinflammation, and underlies extracellular matrix proteolytic dysfunction at the neurovascular interface. All these pathological pathways and interactions worsen BBB breakdown and hemorrhagic transformation after tPA thrombolytic therapy in DM stroke.

by activated leukocytes is one of key pathological factors contributing to BBB leakage and hemorrhagic transformation in ischemic stroke (Lo et al., 2002; Wang and Lo, 2003; Lee et al., 2004; Bao Dang et al., 2013). Importantly, activated extracellular proteases such as MMP-9 act as inflammatory mediators as well, for example, by triggering cytokine expression (Radisky et al., 2005; Amantea et al., 2007; Bao Dang et al., 2013).

In the context of extracellular proteolytic dysregulation in tPA stroke thrombolysis-related hemorrhagic complication, the primary focus is based on the interaction between tPA and MMP-9 (Adibhatla and Hatcher, 2008; Wang et al., 2008; Jin et al., 2010; Lakhan et al., 2013). tPA amplifies MMP-9 expression, which may increase oxidative stress and lead to neuroinflammation (Lo et al., 2004; Wang et al., 2004), (Wang et al., 2003; Zhang et al., 2007). Experimental data suggest that extracellular matrix proteolysis may target multiple cell types at the neurovascular interface, and underlie multiple cascades of BBB disruption after tPA reperfusion treatment (Lee et al., 2004; Lo et al., 2004; Seo et al., 2012).

Fundamentally, the precise pathological molecular mechanisms involved in DM induced complications after stroke remain poorly characterized (Desilles et al., 2013; Hafez et al., 2014), especially because assessing dynamic changes and interactions in multiple factors is technically difficult, particularly in *in vivo* animal models. As discussed earlier, because there are multiple upstream regulators of neurovascular proteolysis-BBB disruption comprising mediators of hyperglycemia, oxidative stress, and inflammation, exogenous tPA may contribute to this pathological process (Kaur et al., 2004; Lo et al., 2004; Adibhatla and Hatcher, 2008; Wang et al., 2008; Jin et al., 2010; Fan et al., 2014). Therefore, in

combination with tPA in patients with diabetes/post stroke hyperglycemia, targeting these upstream mechanisms, or even multiple combinations of these mechanisms may ultimately improve stroke thrombolytic therapy in both safety and efficacy (Fan et al., 2014; Kanazawa et al., 2017; Knecht et al., 2018).

Summary

DM is recognized as a clinically important vascular comorbidity that leads to lower recanalization rates and increased risks of hemorrhagic transformation. In this mini-review, we summarized recent advances in the understanding of the underlying mechanisms involved. Potential pathological factors that relate to suboptimal recanalization include higher plasma PAI-1, diabetic atherogenic vascular damage, glycation of the tPA receptor annexin A2, alterations in fibrin clot density, and impaired collaterals (Figure 1). Factors that may contribute to hemorrhagic transformation include hyperglycemia, vascular oxidative stress, inflammation, tPA neurovascular toxicity, and dysfunction in extracellular proteolysis balance (Figure 2). A better understanding of these complex pathways may eventually lead to novel ways of counteracting the negative effects of DM in stroke.

The conflicts of interest statement

The authors declare that they have no conflicts of interest.

References

- Adams HP, Jr., del Zoppo G, Alberts MJ, Bhatt DL, Brass L, Furlan A, Grubb RL, Higashida RT, Jauch EC, Kidwell C, Lyden PD, Morgenstern LB, Qureshi AI, Rosenwasser RH, Scott PA, Wijndicks EF (2007) Guidelines for the early management of adults with ischemic stroke: a guideline from the American Heart Association/American Stroke Association Stroke Council, Clinical Cardiology Council, Cardiovascular Radiology and Intervention Council, and the Atherosclerotic Peripheral Vascular Disease and Quality of Care Outcomes in Research Interdisciplinary Working Groups: The American Academy of Neurology affirms the value of this guideline as an educational tool for neurologists. *Circulation* 115:e478-534.
- Adibhatla RM, Hatcher JF (2008) Tissue plasminogen activator (tPA) and matrix metalloproteinases in the pathogenesis of stroke: therapeutic strategies. *CNS Neurol Disord Drug Targets* 7:243-253.
- Ahmed N, Davalos A, Eriksson N, Ford GA, Glahn J, Hennerici M, Mikulik R, Kaste M, Lees KR, Lindsberg PJ, Toni D (2010) Association of admission blood glucose and outcome in patients treated with intravenous thrombolysis: results from the Safe Implementation of Treatments in Stroke International Stroke Thrombolysis Register (SITS-ISTR). *Archives of neurology* 67:1123-1130.
- Air EL, Kissela BM (2007) Diabetes, the metabolic syndrome, and ischemic stroke: epidemiology and possible mechanisms. *Diabetes Care* 30:3131-3140.
- Akamatsu Y, Nishijima Y, Lee CC, Yang SY, Shi L, An L, Wang RK, Tominaga T, Liu J (2015) Impaired leptomeningeal collateral flow contributes to the poor outcome following experimental stroke in the Type 2 diabetic mice. *J Neurosci* 35:3851-3864.
- Alessi MC, Poggi M, Juhan-Vague I (2007) Plasminogen activator inhibitor-1, adipose tissue and insulin resistance. *Current opinion in lipidology* 18:240-245.
- Alexandrov AV, Grotta JC (2002) Arterial reocclusion in stroke patients treated with intravenous tissue plasminogen activator. *Neurology* 59:862-867.
- Allport L, Baird T, Butcher K, Macgregor L, Prosser J, Colman P, Davis S (2006) Frequency and temporal profile of poststroke hyperglycemia using continuous glucose monitoring. *Diabetes Care* 29:1839-1844.
- Alzahrani SH, Ajjan RA (2010) Coagulation and fibrinolysis in diabetes. *Diab Vasc Dis Res* 7:260-273.
- Amantea D, Russo R, Gliozzi M, Fratto V, Berliocchi L, Bagetta G, Bernardi G, Corasaniti MT (2007) Early upregulation of matrix metalloproteinases following reperfusion triggers neuroinflammatory mediators in brain ischemia in rat. *Int Rev Neurobiol* 82:149-169.
- Amantea D, Tassorelli C, Petrelli F, Certo M, Bezzi P, Micieli G, Corasaniti MT, Bagetta G (2013) Understanding the Multifaceted Role of Inflammatory Mediators in Ischemic Stroke. *Curr Med Chem*.
- Asahi M, Asahi K, Wang X, Lo EH (2000) Reduction of tissue plasminogen activator-induced hemorrhage and brain injury by free radical spin trapping after embolic focal cerebral ischemia in rats. *J Cereb Blood Flow Metab* 20:452-457.
- Aso Y (2007) Plasminogen activator inhibitor (PAI)-1 in vascular inflammation and thrombosis. *Front Biosci* 12:2957-2966.
- Bambauer KZ, Johnston SC, Bambauer DE, Zivin JA (2006) Reasons why few patients with acute stroke receive tissue plasminogen activator. *Arch Neurol* 63:661-664.
- Bao Dang Q, Lapergue B, Tran-Dinh A, Diallo D, Moreno JA, Mazighi M, Romero IA, Weksler B, Michel JB, Amarenco P, Meilhac O (2013) High-density lipoproteins limit neutrophil-induced damage to the blood-brain barrier *in vitro*. *J Cereb Blood Flow Metab* 33:575-582.
- Beckman JA, Paneni F, Cosentino F, Creager MA (2013) Diabetes and vascular disease: pathophysiology, clinical consequences, and medical therapy: part II. *Eur Heart J* 34:2444-2452.
- Borlongan CV, Glover LE, Sanberg PR, Hess DC (2012) Permeating the blood brain barrier and abrogating the inflammation in stroke: implications for stroke therapy. *Curr Pharm Des* 18:3670-3676.
- Bruno A, Kent TA, Coull BM, Shankar RR, Saha C, Becker KJ, Kissela BM, Williams LS (2008) Treatment of hyperglycemia in ischemic stroke (THIS): a randomized pilot trial. *Stroke* 39:384-389.
- Bruno A, Levine SR, Frankel MR, Brott TG, Lin Y, Tilley BC, Lyden PD, Broderick JP, Kwiatkowski TG, Fineberg SE (2002) Admission glucose level and clinical outcomes in the NINDS rt-PA Stroke Trial. *Neurology* 59:669-674.
- Chapman SN, Mehndiratta P, Johansen MC, McMurphy TL, Johnston KC, Southerland AM (2014) Current perspectives on the use of intravenous recombinant tissue plasminogen activator (tPA) for treatment of acute ischemic stroke. *Vasc Health Risk Manag* 10:75-87.
- Chen R, Ovbiagele B, Feng W (2016) Diabetes and Stroke: Epidemiology, Pathophysiology, Pharmaceuticals and Outcomes. *Am J Med Sci* 351:380-386.
- Cronin CA (2010) Intravenous tissue plasminogen activator for stroke: a review of the ECASS III results in relation to prior clinical trials. *J Emerg Med* 38:99-105.
- Dai H, Yu Z, Fan X, Liu N, Yan M, Chen Z, Lo EH, Hajjar KA, Wang X (2013) Dysfunction of annexin A2 contributes to hyperglycaemia-induced loss of human endothelial cell surface fibrinolytic activity. *Thromb Haemost* 109:1070-1078.
- Davalos A (2005) Thrombolysis in acute ischemic stroke: successes, failures, and new hopes. *Cerebrovasc Dis* 20 Suppl 2:135-139.
- Desilles JP, Meseguer E, Labreuche J, Lapergue B, Sirimarco G, Gonzalez-Valcarcel J, Lavallee P, Cabrejo L, Guidoux C, Klein I, Amarenco P, Mazighi M (2013) Diabetes mellitus, admission glucose, and outcomes after stroke thrombolysis: a registry and systematic review. *Stroke*

- 44:1915-1923.
- Dunn EJ, Ariens RA, Grant PJ (2005) The influence of type 2 diabetes on fibrin structure and function. *Diabetologia* 48:1198-1206.
- Dunn EJ, Philippou H, Ariens RA, Grant PJ (2006) Molecular mechanisms involved in the resistance of fibrin to clot lysis by plasmin in subjects with type 2 diabetes mellitus. *Diabetologia* 49:1071-1080.
- Fagan SC, Hess DC, Hohnadel EJ, Pollock DM, Ergul A (2004) Targets for vascular protection after acute ischemic stroke. *Stroke* 35:2220-2225.
- Faigle R, Sharrief A, Marsh EB, Llinas RH, Urrutia VC (2014) Predictors of critical care needs after IV thrombolysis for acute ischemic stroke. *PLoS One* 9:e88652.
- Fan X, Jiang Y, Yu Z, Yuan J, Sun X, Xiang S, Lo EH, Wang X (2014) Combination approaches to attenuate hemorrhagic transformation after tPA thrombolytic therapy in patients with poststroke hyperglycemia/diabetes. *Adv Pharmacol* 71:391-410.
- Fan X, Jiang Y, Yu Z, Liu Q, Guo S, Sun X, van Leyen K, Ning M, Gao X, Lo EH, Wang X (2017) Annexin A2 Plus Low-Dose Tissue Plasminogen Activator Combination Attenuates Cerebrovascular Dysfunction After Focal Embolic Stroke of Rats. *Transl Stroke Res* 8:549-559.
- Gasche Y, Copin JC, Sugawara T, Fujimura M, Chan PH (2001) Matrix metalloproteinase inhibition prevents oxidative stress-associated blood-brain barrier disruption after transient focal cerebral ischemia. *J Cereb Blood Flow Metab* 21:1393-1400.
- Ghitescu LD, Gugliucci A, Dumas F (2001) Actin and annexins I and II are among the main endothelial plasmalemma-associated proteins forming early glucose adducts in experimental diabetes. *Diabetes* 50:1666-1674.
- Gidday JM, Gasche YG, Copin JC, Shah AR, Perez RS, Shapiro SD, Chan PH, Park TS (2005) Leukocyte-derived matrix metalloproteinase-9 mediates blood-brain barrier breakdown and is proinflammatory after transient focal cerebral ischemia. *Am J Physiol Heart Circ Physiol* 289:H558-568.
- Gugliucci A, Ghitescu L (2002) Is diabetic hypercoagulability an acquired annexinopathy? Glycation of annexin II as a putative mechanism for impaired fibrinolysis in diabetic patients. *Medical hypotheses* 59:247-251.
- Hafez S, Coucha M, Bruno A, Fagan SC, Ergul A (2014) Hyperglycemia, Acute Ischemic Stroke, and Thrombolytic Therapy. *Transl Stroke Res*.
- Harsany M, Tsvigoulis G, Alexandrov AV (2014) Intravenous thrombolysis in acute ischemic stroke: standard and potential future applications. *Expert Rev Neurother* 14:879-892.
- Hill MD (2014) Stroke and diabetes mellitus. *Handb Clin Neurol* 126:167-174.
- Ishrat T, Soliman S, Guan W, Saler M, Fagan SC (2012) Vascular protection to increase the safety of tissue plasminogen activator for stroke. *Curr Pharm Des* 18:3677-3684.
- Jauch EC, Saver JL, Adams HP, Jr., Bruno A, Connors JJ, Demaerschalk BM, Khatri P, McMullan PW, Jr., Qureshi AI, Rosenfield K, Scott PA, Summers DR, Wang DZ, Wintermark M, Yonas H, American Heart Association Stroke C, Council on Cardiovascular N, Council on Peripheral Vascular D, Council on Clinical C (2013) Guidelines for the early management of patients with acute ischemic stroke: a guideline for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke* 44:870-947.
- Jian Liu K, Rosenberg GA (2005) Matrix metalloproteinases and free radicals in cerebral ischemia. *Free Radic Biol Med* 39:71-80.
- Jiang Y, Fan X, Yu Z, Liao Z, Wang XS, van Leyen K, Sun X, Lo EH, Wang X (2015) Combination Low-Dose Tissue-Type Plasminogen Activator Plus Annexin A2 for Improving Thrombolytic Stroke Therapy. *Front Cell Neurosci* 9:397.
- Jin R, Yang G, Li G (2010) Molecular insights and therapeutic targets for blood-brain barrier disruption in ischemic stroke: critical role of matrix metalloproteinases and tissue-type plasminogen activator. *Neurobiol Dis* 38:376-385.
- Kanazawa M, Takahashi T, Nishizawa M, Shimohata T (2017) Therapeutic Strategies to Attenuate Hemorrhagic Transformation After Tissue Plasminogen Activator Treatment for Acute Ischemic Stroke. *J Atheroscler Thromb* 24:240-253.
- Kase CS, Furlan AJ, Wechsler LR, Higashida RT, Rowley HA, Hart RG, Molinari GF, Frederick LS, Roberts HC, Gebel JM, Sila CA, Schulz GA, Roberts RS, Gent M (2001) Cerebral hemorrhage after intra-arterial thrombolysis for ischemic stroke: the PROACT II trial. *Neurology* 57:1603-1610.
- Kaur J, Zhao Z, Klein GM, Lo EH, Buchan AM (2004) The neurotoxicity of tissue plasminogen activator? *J Cereb Blood Flow Metab* 24:945-963.
- Kelly PJ, Morrow JD, Ning M, Koroshetz W, Lo EH, Terry E, Milne GL, Hubbard J, Lee H, Stevenson E, Lederer M, Furie KL (2008) Oxidative stress and matrix metalloproteinase-9 in acute ischemic stroke: the Biomarker Evaluation for Antioxidant Therapies in Stroke (BEAT-Stroke) study. *Stroke* 39:100-104.
- Kimura K, Iguchi Y, Shibasaki K, Iwanaga T, Aoki J (2009) Recanalization of the MCA should play an important role in dramatic recovery after t-PA therapy in patients with ICA occlusion. *J Neurol Sci* 285:130-133.
- Knecht T, Borlongan C, Dela Pena I (2018) Combination therapy for ischemic stroke: Novel approaches to lengthen therapeutic window of tissue plasminogen activator. *Brain Circ* 4:99-108.
- Konieczynska M, Fil K, Bazanek M, Undas A (2014) Prolonged duration of type 2 diabetes is associated with increased thrombin generation, prothrombotic fibrin clot phenotype and impaired fibrinolysis. *Thromb Haemost* 111:685-693.
- Kruyt ND, Biessels GJ, Devries JH, Roos YB (2010) Hyperglycemia in acute ischemic stroke: pathophysiology and clinical management. *Nat Rev Neurol* 6:145-155.
- Kwon JH, Kwon SU, Lee JH, Choi CG, Suh DC, Kim JS (2004) Factors affecting the angiographic recanalization and early clinical improvement in middle cerebral artery territory infarction after thrombolysis. *Arch Neurol* 61:1682-1686.
- Lakhan SE, Kirchgessner A, Tepper D, Leonard A (2013) Matrix metalloproteinases and blood-brain barrier disruption in acute ischemic stroke. *Front Neurol* 4:32.
- Lapchak PA, Chapman DF, Zivin JA (2001) Pharmacological effects of the spin trap agents N-t-butyl-phenylnitron (PBN) and 2,2,6,6-tetramethylpiperidine-N-oxyl (TEMPO) in a rabbit thromboembolic stroke model: combination studies with the thrombolytic tissue plasminogen activator. *Stroke* 32:147-153.
- Lapchak PA, Araujo DM, Song D, Wei J, Purdy R, Zivin JA (2002) Effects of the spin trap agent disodium- [tert-butylimino)methyl]benzene-1,3-disulfonate N-oxide (generic NXY-059) on intracerebral hemorrhage in a rabbit Large clot embolic stroke model: combination studies with tissue plasminogen activator. *Stroke* 33:1665-1670.
- Lee SR, Wang X, Tsuji K, Lo EH (2004) Extracellular proteolytic pathophysiology in the neurovascular unit

- after stroke. *Neurol Res* 26:854-861.
- Lemkes BA, Hermanides J, Devries JH, Holleman F, Meijers JC, Hoekstra JB (2010) Hyperglycemia: a prothrombotic factor? *J Thromb Haemost* 8:1663-1669.
- Lenart N, Brough D, Denes A (2016) Inflammation link vascular disease with neuroinflammation and brain disorders. *J Cereb Blood Flow Metab* 36:1668-1685.
- Li W, Ward R, Valenzuela JP, Dong G, Fagan SC, Ergul A (2017) Diabetes Worsens Functional Outcomes in Young Female Rats: Comparison of Stroke Models, Tissue Plasminogen Activator Effects, and Sexes. *Transl Stroke Res*.
- Liebeskind DS, Tomsick TA, Foster LD, Yeatts SD, Carrozzella J, Demchuk AM, Jovin TG, Khatri P, von Kummer R, Sugg RM, Zaidat OO, Hussain SI, Goyal M, Menon BK, Al Ali F, Yan B, Palesch YY, Broderick JP, Investigators II (2014) Collaterals at angiography and outcomes in the Interventional Management of Stroke (IMS) III trial. *Stroke* 45:759-764.
- Linfante I, Llinas RH, Selim M, Chaves C, Kumar S, Parker RA, Caplan LR, Schlaug G (2002) Clinical and vascular outcome in internal carotid artery versus middle cerebral artery occlusions after intravenous tissue plasminogen activator. *Stroke* 33:2066-2071.
- Ling Q, Jacovina AT, Deora A, Febbraio M, Simantov R, Silverstein RL, Hempstead B, Mark WH, Hajjar KA (2004) Annexin II regulates fibrin homeostasis and neoangiogenesis *in vivo*. *J Clin Invest* 113:38-48.
- Lo EH, Wang X, Cuzner ML (2002) Extracellular proteolysis in brain injury and inflammation: role for plasminogen activators and matrix metalloproteinases. *J Neurosci Res* 69:1-9.
- Lo EH, Broderick JP, Moskowitz MA (2004) tPA and proteolysis in the neurovascular unit. *Stroke* 35:354-356.
- Ly H, Verma N, Wu F, Liu M, Saatman KE, Nelson PT, Slevin JT, Goldstein LB, Biessels GJ, Despa F (2017) Brain microvascular injury and white matter disease provoked by diabetes-associated hyperamylinemia. *Ann Neurol* 82:208-222.
- Maatman BT, Schmeisser G, Kreutz RP (2018) Fibrin Clot Strength in Patients with Diabetes Mellitus Measured by Thrombelastography. *J Diabetes Res* 2018:4543065.
- Manning NW, Campbell BC, Oxley TJ, Chapot R (2014) Acute ischemic stroke: time, penumbra, and reperfusion. *Stroke* 45:640-644.
- Miller DJ, Simpson JR, Silver B (2011) Safety of thrombolysis in acute ischemic stroke: a review of complications, risk factors, and newer technologies. *Neurohospitalist* 1:138-147.
- Molina CA, Alexandrov AV, Demchuk AM, Saqqur M, Uchino K, Alvarez-Sabin J, Investigators C (2004) Improving the predictive accuracy of recanalization on stroke outcome in patients treated with tissue plasminogen activator. *Stroke* 35:151-156.
- Molina CA, Montaner J, Abilleira S, Arenillas JF, Ribo M, Huertas R, Romero F, Alvarez-Sabin J (2001) Time course of tissue plasminogen activator-induced recanalization in acute cardioembolic stroke: a case-control study. *Stroke* 32:2821-2827.
- Montaner J (2009) Blood biomarkers to guide stroke thrombolysis. *Front Biosci (Elite Ed)* 1:200-208.
- Najjar S, Pearlman DM, Devinsky O, Najjar A, Zagzag D (2013) Neurovascular unit dysfunction with blood-brain barrier hyperpermeability contributes to major depressive disorder: a review of clinical and experimental evidence. *J Neuroinflammation* 10:142.
- Nishijima Y, Akamatsu Y, Yang SY, Lee CC, Baran U, Song S, Wang RK, Tominaga T, Liu J (2016) Impaired Collateral Flow Compensation During Chronic Cerebral Hypoperfusion in the Type 2 Diabetic Mice. *Stroke* 47:3014-3021.
- Nogueira RG et al. (2018) Thrombectomy 6 to 24 Hours after Stroke with a Mismatch between Deficit and Infarct. *N Engl J Med* 378:11-21.
- Pandolfi A, Giaccari A, Cilli C, Alberta MM, Morviducci L, De Filippis EA, Buongiorno A, Pellegrini G, Capani F, Consoli A (2001) Acute hyperglycemia and acute hyperinsulinemia decrease plasma fibrinolytic activity and increase plasminogen activator inhibitor type 1 in the rat. *Acta Diabetol* 38:71-76.
- Picanco MR, Christensen S, Campbell BC, Churilov L, Parsons MW, Desmond PM, Barber PA, Levi CR, Bladin CF, Donnan GA, Davis SM, Investigators E (2014) Reperfusion after 4.5 hours reduces infarct growth and improves clinical outcomes. *Int J Stroke* 9:266-269.
- Poppe AY, Majumdar SR, Jeerakathil T, Ghali W, Buchan AM, Hill MD (2009) Admission hyperglycemia predicts a worse outcome in stroke patients treated with intravenous thrombolysis. *Diabetes Care* 32:617-622.
- Pun PB, Lu J, Mochhala S (2009) Involvement of ROS in BBB dysfunction. *Free Radic Res* 43:348-364.
- Radisky DC, Levy DD, Littlepage LE, Liu H, Nelson CM, Fata JE, Leake D, Godden EL, Albertson DG, Nieto MA, Werb Z, Bissell MJ (2005) Rac1b and reactive oxygen species mediate MMP-3-induced EMT and genomic instability. *Nature* 436:123-127.
- Reiter M, Teuschl Y, Matz K, Seyfang L, Brainin M, Austrian Stroke Unit Registry C (2014) Diabetes and thrombolysis for acute stroke: a clear benefit for diabetics. *Eur J Neurol* 21:5-10.
- Ribo M, Montaner J, Molina CA, Arenillas JF, Santamarina E, Alvarez-Sabin J (2004a) Admission fibrinolytic profile predicts clot lysis resistance in stroke patients treated with tissue plasminogen activator. *Thromb Haemost* 91:1146-1151.
- Ribo M, Montaner J, Molina CA, Arenillas JF, Santamarina E, Quintana M, Alvarez-Sabin J (2004b) Admission fibrinolytic profile is associated with symptomatic hemorrhagic transformation in stroke patients treated with tissue plasminogen activator. *Stroke; a journal of cerebral circulation* 35:2123-2127.
- Ribo M, Molina C, Montaner J, Rubiera M, Delgado-Mederos R, Arenillas JF, Quintana M, Alvarez-Sabin J (2005) Acute hyperglycemia state is associated with lower tPA-induced recanalization rates in stroke patients. *Stroke* 36:1705-1709.
- Rosenberg GA (2002) Matrix metalloproteinases in neuroinflammation. *Glia* 39:279-291.
- Rubiera M, Alvarez-Sabin J, Ribo M, Montaner J, Santamarina E, Arenillas JF, Huertas R, Delgado P, Purroy F, Molina CA (2005) Predictors of early arterial recanalization after tissue plasminogen activator-induced recanalization in acute ischemic stroke. *Stroke* 36:1452-1456.
- Seet RC, Rabinstein AA (2012) Symptomatic intracranial hemorrhage following intravenous thrombolysis for acute ischemic stroke: a critical review of case definitions. *Cerebrovasc Dis* 34:106-114.
- Seo JH, Guo S, Lok J, Navaratna D, Whalen MJ, Kim KW, Lo EH (2012) Neurovascular matrix metalloproteinases and the blood-brain barrier. *Curr Pharm Des* 18:3645-3648.
- Shrestha S, Poudel RS, Thapa LJ, Khatiwada D (2014) Intravenous Thrombolysis and Risk Factors for Ischemic Stroke. *JNMA J Nepal Med Assoc* 52:745-750.
- Simi A, Tsakiri N, Wang P, Rothwell NJ (2007) Interleukin-1 and inflammatory neurodegeneration. *Biochem Soc Trans* 35:1122-1126.
- Tang H, Zhang S, Yan S, Liebeskind DS, Sun J, Ding X, Zhang

- M, Lou M (2016) Unfavorable neurological outcome in diabetic patients with acute ischemic stroke is associated with incomplete recanalization after intravenous thrombolysis. *J Neurointerv Surg* 8:342-346.
- Thomassen L, Bakke SJ (2007) Endovascular reperfusion therapy in acute ischaemic stroke. *Acta Neurol Scand Suppl* 187:22-29.
- Tjarnlund-Wolf A, Brogren H, Lo EH, Wang X (2012) Plasminogen activator inhibitor-1 and thrombotic cerebrovascular diseases. *Stroke* 43:2833-2839.
- Tureyen K, Bowen K, Liang J, Dempsey RJ, Vemuganti R (2011) Exacerbated brain damage, edema and inflammation in type-2 diabetic mice subjected to focal ischemia. *J Neurochem* 116:499-507.
- Vaidyula VR, Rao AK, Mozzoli M, Homko C, Cheung P, Boden G (2006) Effects of hyperglycemia and hyperinsulinemia on circulating tissue factor procoagulant activity and platelet CD40 ligand. *Diabetes* 55:202-208.
- Venkat P, Chopp M, Chen J (2017) Blood-Brain Barrier Disruption, Vascular Impairment, and Ischemia/Reperfusion Damage in Diabetic Stroke. *J Am Heart Assoc* 6.
- Walter T, Szabo S, Suselbeck T, Borggreffe M, Lang S, Swoboda S, Hoffmeister HM, Dempfle CE (2010) Effect of atorvastatin on haemostasis, fibrinolysis and inflammation in normocholesterolaemic patients with coronary artery disease: a post hoc analysis of data from a prospective, randomized, double-blind study. *Clinical drug investigation* 30:453-460.
- Wang X, Lo EH (2003) Triggers and mediators of hemorrhagic transformation in cerebral ischemia. *Mol Neurobiol* 28:229-244.
- Wang X, Rosell A, Lo EH (2008) Targeting extracellular matrix proteolysis for hemorrhagic complications of tPA stroke therapy. *CNS Neurol Disord Drug Targets* 7:235-242.
- Wang X, Lee SR, Arai K, Lee SR, Tsuji K, Rebeck GW, Lo EH (2003) Lipoprotein receptor-mediated induction of matrix metalloproteinase by tissue plasminogen activator. *Nat Med* 9:1313-1317.
- Wang X, Tsuji K, Lee SR, Ning M, Furie KL, Buchan AM, Lo EH (2004) Mechanisms of hemorrhagic transformation after tissue plasminogen activator reperfusion therapy for ischemic stroke. *Stroke* 35:2726-2730.
- Wang X, Fan X, Yu Z, Liao Z, Zhao J, Mandeville E, Guo S, Lo EH, Wang X (2014) Effects of tissue plasminogen activator and annexin A2 combination therapy on long-term neurological outcomes of rat focal embolic stroke. *Stroke* 45:619-622.
- Weintraub MI (2006) Thrombolysis (tissue plasminogen activator) in stroke: a medicolegal quagmire. *Stroke* 37:1917-1922.
- Whiteley W (2011) Identifying blood biomarkers to improve the diagnosis of stroke. *The journal of the Royal College of Physicians of Edinburgh* 41:152-154.
- Whiteley WN, Thompson D, Murray G, Cohen G, Lindley RI, Wardlaw J, Sandercock P (2014) Targeting recombinant tissue-type plasminogen activator in acute ischemic stroke based on risk of intracranial hemorrhage or poor functional outcome: an analysis of the third international stroke trial. *Stroke* 45:1000-1006.
- Won SJ, Tang XN, Suh SW, Yenari MA, Swanson RA (2011) Hyperglycemia promotes tissue plasminogen activator-induced hemorrhage by increasing superoxide production. *Ann Neurol* 70:583-590.
- Zhang X, Polavarapu R, She H, Mao Z, Yepes M (2007) Tissue-type plasminogen activator and the low-density lipoprotein receptor-related protein mediate cerebral ischemia-induced nuclear factor-kappaB pathway activation. *Am J Pathol* 171:1281-1290.